



Juniper Pines Poultry Lab

DNA Testing Information as of April 1, 2026

Pricing

- \$8 per test - gender or blue egg

How Samples Work

You collect the blood samples yourself, ideally on a **coffee-filter triangle**. Each sample must be **fully dry**, placed in its **own baggie**, and clearly labeled with **your identifier for that bird**. For feather samples, see Option B.

Payment

~Contact me for *any* questions at [719-285-9686](tel:719-285-9686). **Text works best** due to my lack of cell service.

~Payment must be made before you mail samples. Once Payment is made, I create a ticket for you and watch for your shipment. Mail is collected Monday, Wednesday, and Friday.

~Upon arrival, I begin testing. Results are communicated within 24 - 48 hours.

~Accepted forms: **Venmo (jpPoultryLab, 3165)**, **Credit Cards**, **PayPal**, and **Zelle** (text for contact info).

~If you're along my route, I can also **pick up samples and payments in person**. Watch FB for my travel info.

Mailing Address

Juniper Pines Poultry Lab
306 E Main St, #627
Canon City, CO 81212

How to Collect Blood Spot Samples

What You'll Need

- Toenail clippers
- Alcohol wipes or rubbing alcohol + cotton (NOT BLEACH)
- Disposable gloves (or clean hands)
- Styptic powder or cornstarch
- Coffee filters (best option) or an index card (harder to dry)
- Your chosen chick identifier (leg band, bracelet, pet name, or even "brown chick")
- One baggie **per** sample

Tips:

- Cut coffee filters like a pizza into triangles—hold the wide end and collect the blood on the pointy end.
- It's easier to **write the ID on the filter before** collecting the blood.
- Any ID system is fine as long as **you** know which chick is which and the sample is clearly marked.

Step-by-Step Directions

- **Clean your clippers**
Wipe the cutting edge with an alcohol wipe. (Don't dunk—dunking doesn't remove debris.)
- **Pick up the chick and apply its ID**
Band the leg or note whatever identifier you're using so it matches the sample.
- **Prep the nail**
Wipe one toenail with alcohol and let it dry for a few seconds.
- **Clip & collect**
Clip the nail short, just enough for a small drop of blood to form.
Set the clippers down, pick up your coffee-filter triangle, and touch the tip to the drop.
You only need a spot about the size of a **grain of rice**—anywhere on the paper, as long as it's not smeared or touched. I don't need much, but in case of PCR malfunction, it's best to have more blood available to sample.

Trick: Bend the tip of the filter upward so it stands up while drying.
- **Stop the bleeding**
Dab the chick's nail in cornstarch or styptic powder.
Give the chick a little kiss and return it to the brooder.
- **Clean up & repeat**
Wipe your clippers again, make sure the sample is labeled, and move on to the next chick.
- **Dry & package**
Once **all samples are completely dry**, place each one in its **own** ziplock baggie.
Put all baggies into a mailing envelope along with a sheet of paper containing your **contact information**.
- **Option B** for older birds or other species that you don't feel comfortable clipping toenails (Parrot, Emu, Pigeon, etc.):
Pluck 3-5 feathers from the breast or wing. Ensure the quill bulb (the tiny clear/white bump at the end) is attached. Place feathers in a small envelope and write your Bird ID on the outside. Put all baggies into a mailing envelope along with a sheet of paper containing your **contact information**.

Results are 99% accurate and while I do everything within my power to keep your samples pure, I cannot guarantee errors due to several factors in potential contamination steps - starting at collection. In the case of a failed sample, I will test again with the original sample for free. If I physically cannot see a blood spot on your sample, I will try to test it but might require another submission.